

# Whole-genome mutational landscape and characterization of noncoding and structural mutations in liver cancer

Akihiro Fujimoto<sup>1</sup>, Mayuko Furuta<sup>1</sup>, Yasushi Totoki<sup>2</sup>, Tatsuhiko Tsunoda<sup>1</sup>, Mamoru Kato<sup>2</sup>, Atsushi Ono<sup>4</sup>, Hiroki Yamaue<sup>3</sup>, Kazuaki Chayama<sup>1,4</sup>, Satoru Miyano<sup>5</sup>, Hiroyuki Aburatani<sup>6</sup>, Tatsuhiro Shibata<sup>2</sup>, Hidewaki Nakagawa<sup>1</sup>

(1; IMS, RIKEN, 2; Natl. Cancer Ctr. Res. Inst., 3; Wakayama Med. Univ., 4; Hiroshima Univ. Sch. of Med., 5; Inst. of Med. Sci., The Univ. of Tokyo, 6; RCAST, The Univ. of Tokyo)

# Liver cancer

- ✓ Liver cancer is the third leading cancer-related cause of death and the seventh most common form of cancer in the world.
- ✓ The mortality rate of liver cancer is high in East Asia and Africa, and is also increasing in western countries.
- ✓ One of the major risk factors is virus infection;
  - Hepatitis B virus (HBV):** Asia, Africa
  - Hepatitis C virus (HCV):** Japan (>70% of HCC)
- ✓ Large-scale whole genome sequencing study has not yet been performed for liver cancer.

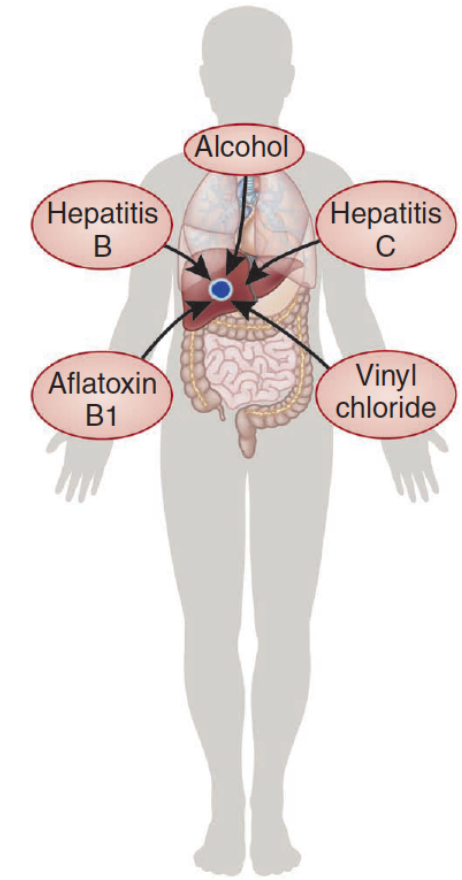


Figure from Zhang *Nat. Genet.* **44**, 1075-7 (2012)

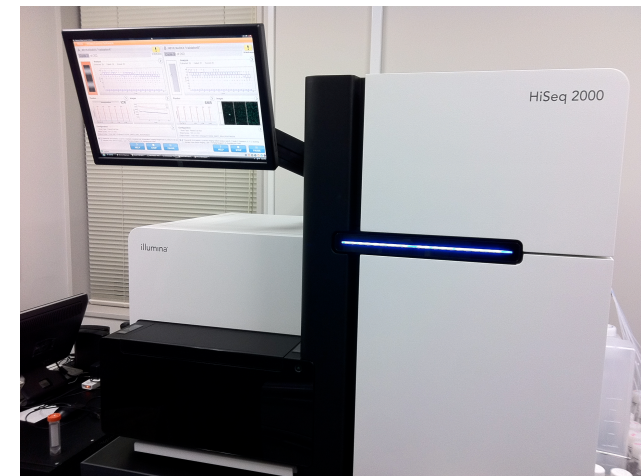
# Topics

- Whole genome sequencing of 300 liver cancer genomes  
Fujimoto *et al.* **Nature Genetics** (2016)
- Circulating tumor DNA analysis for liver cancers  
Ono *et al.* **Cellular and Molecular Gastroenterology and Hepatology** (2015)

Whole genome sequencing of 300 liver cancer genomes  
Fujimoto *et al.* Nature Genetics (2016)

# Sample and sequencing

- **Sample**
  - 300 liver cancers
  - 268 hepatocellular carcinomas (HCCs), 24 intrahepatic cholangiocarcinomas (ICCs), 8 cHCC/ICCs (combined type)
  - 159 HCV, 82 HBV, 55 NBNC and 4 HBV/HCV
  - Whole genome sequencing (WGS) and RNA-sequencing
- **Sequencing**
  - HiSeq2000
  - Paired-end method
  - Library size; 500bp
  - Read length; 100bp



# Somatic mutation calling

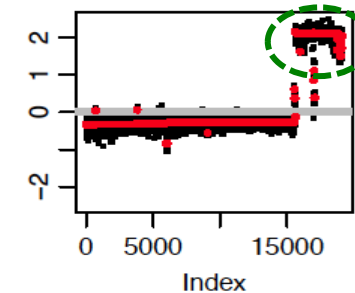
## Point mutation and short indels

- Based on number and frequency of mutant alleles
- False positive rate of point mutation < 3 % in CDS region
- False positive rate of indel < 10 %

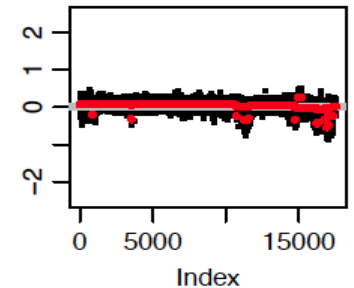
## Copy number alternation

- Ratio of depth of coverage was analyzed by DNACopy<sup>1</sup>

Amplification

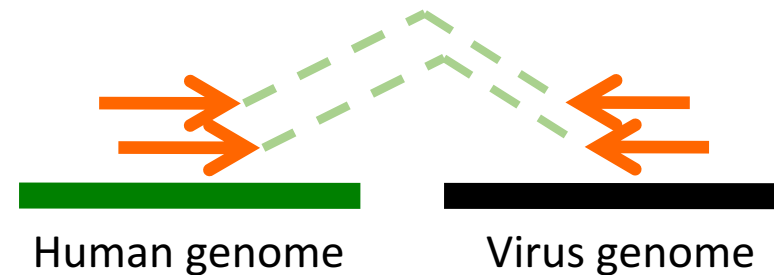
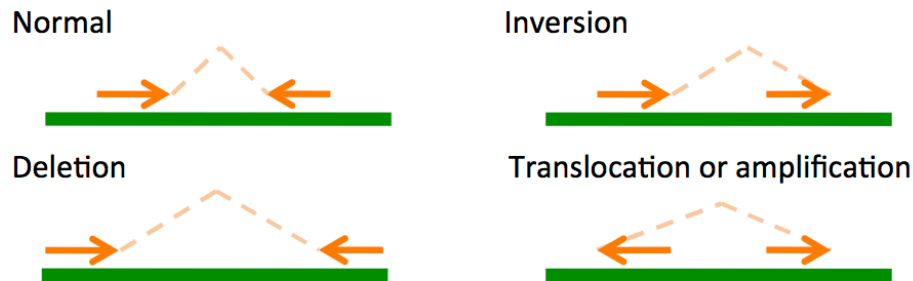


chromosome 14



## Structural variations (STV) and virus integration

- Based on orientation and distance between read-pairs
- Realignment and comparison with other normal samples were performed to exclude false positives



# Contents

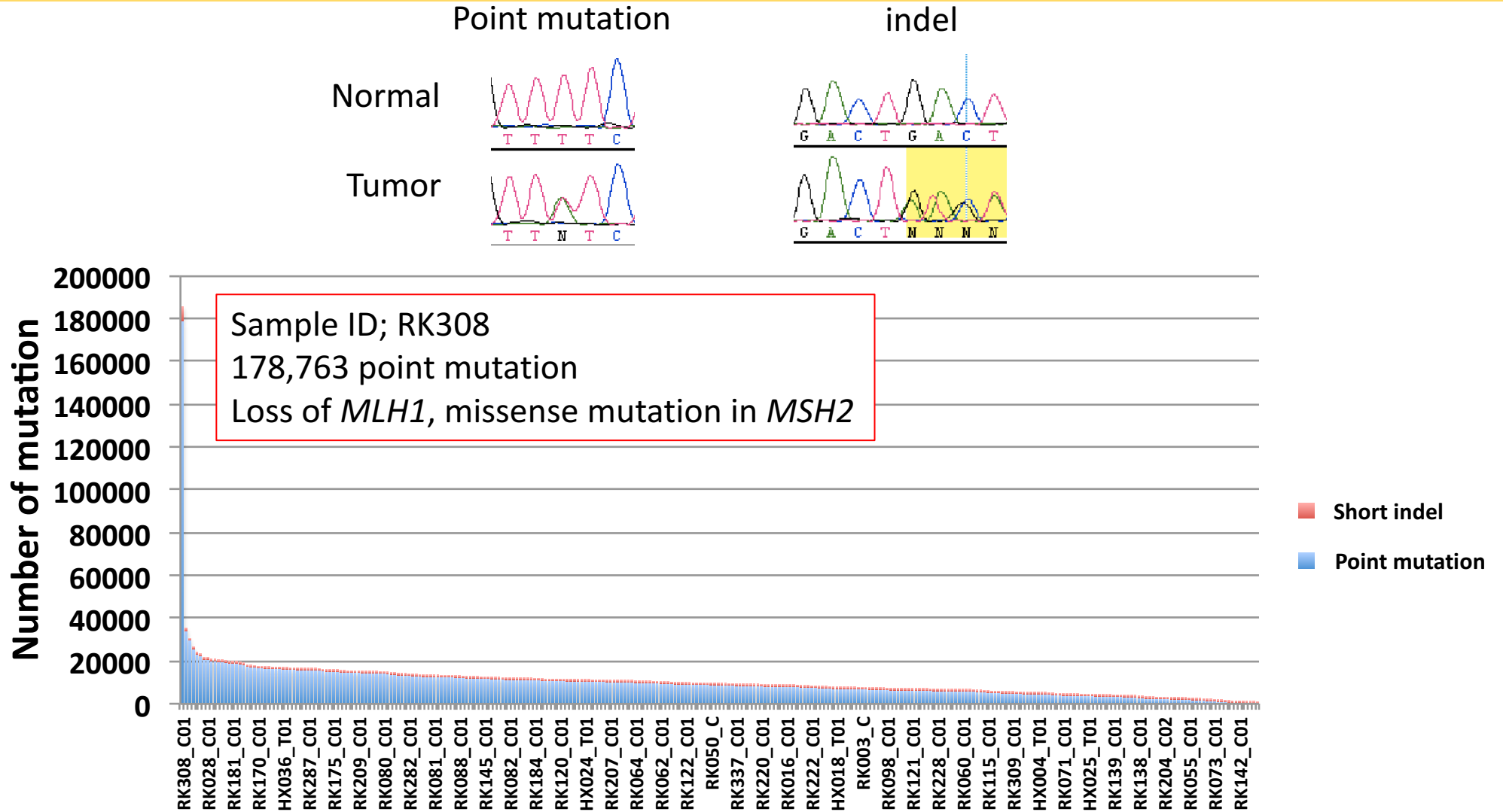
- Pattern of mutation
  - Point mutation and short indel
  - Structural variation
  - Integration of virus into human genome
- Driver genes and noncoding driver regions
  - Significantly mutated genes
  - Significantly mutated noncoding regions
- Structural variations
  - Structural variations
  - Structural variations and gene expression
  - Mutation and gene expression in *TERT*

# Contents

- Pattern of mutation
  - Point mutation and short indel
  - Structural variation
  - Integration of virus into human genome
- Driver genes and noncoding driver regions
  - Significantly mutated genes
  - Significantly mutated noncoding regions
- Structural variations
  - Structural variations
  - Structural variations and gene expression
  - Mutation and gene expression in *TERT*

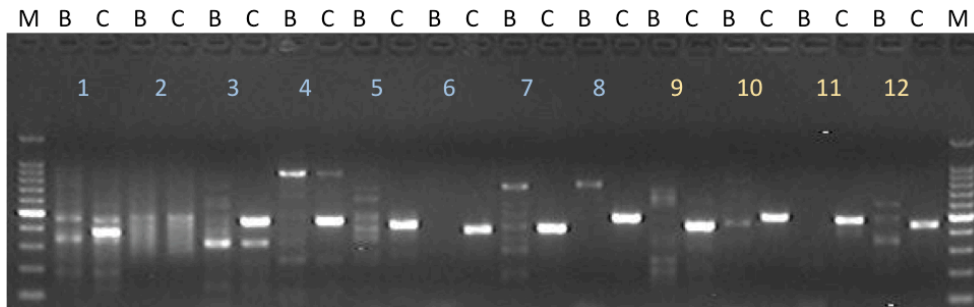


# Point mutation and short indel

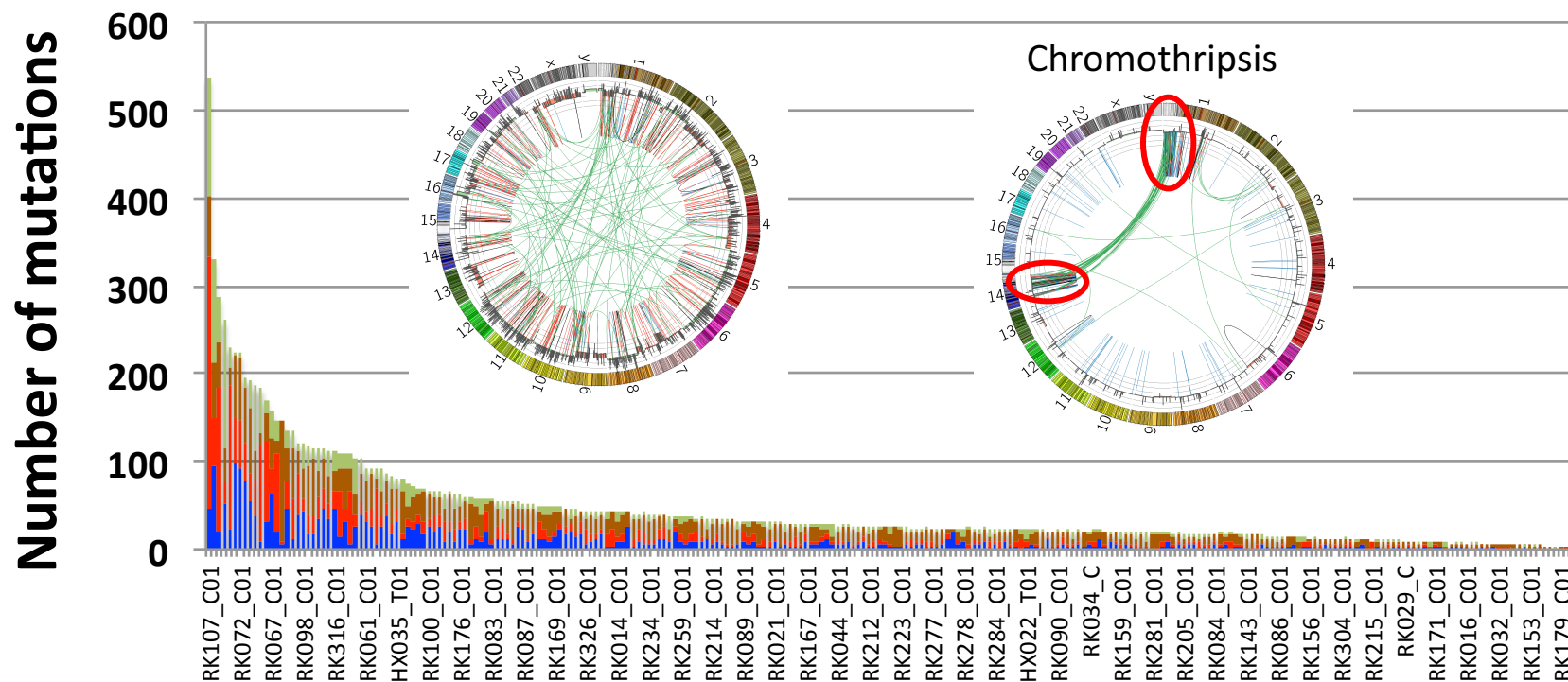


Number of point mutation; 8,777, short indel; 239 (median)

# Structural variation (STV)



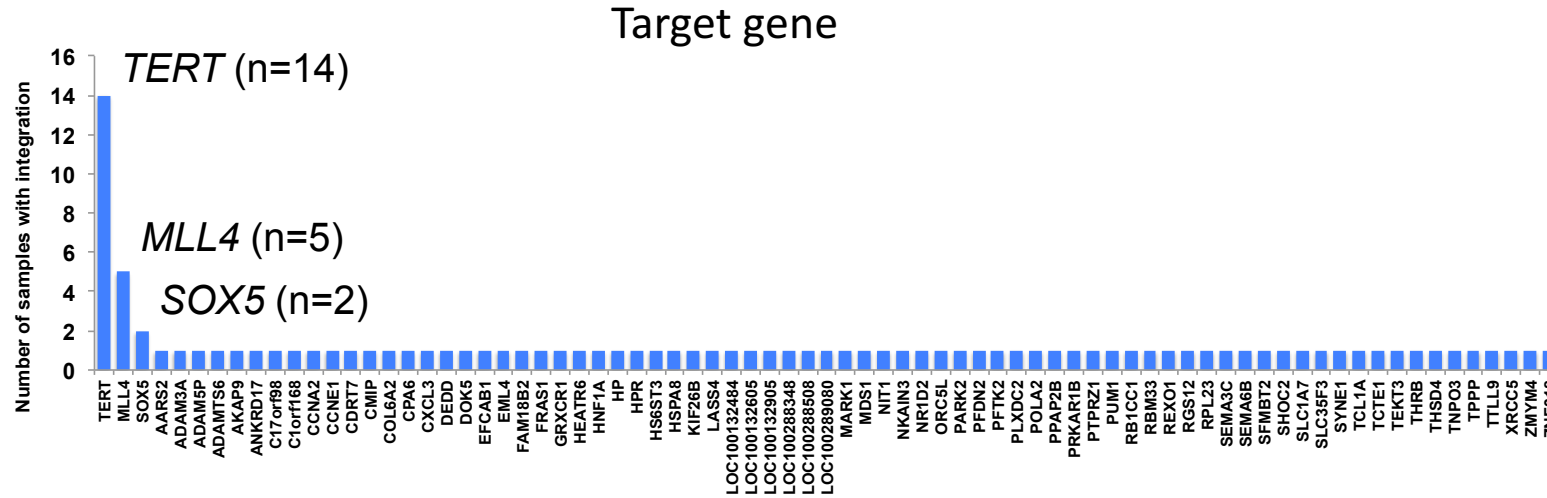
**Inversion** **Tandem duplication** **Deletion** **Chromosomal translocation**



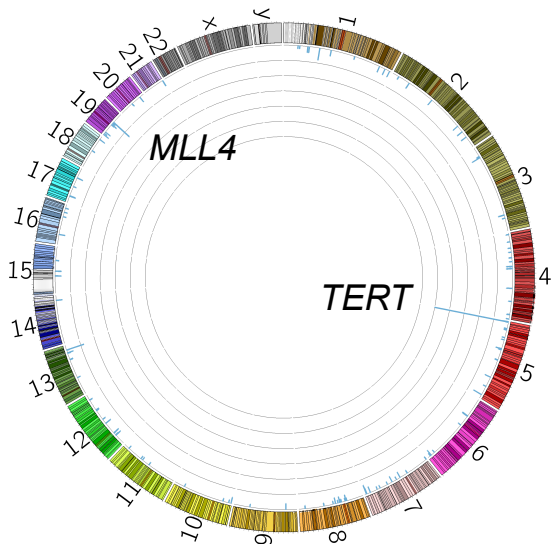
Number of structural variation; 40.6

# Target gene of HBV integration

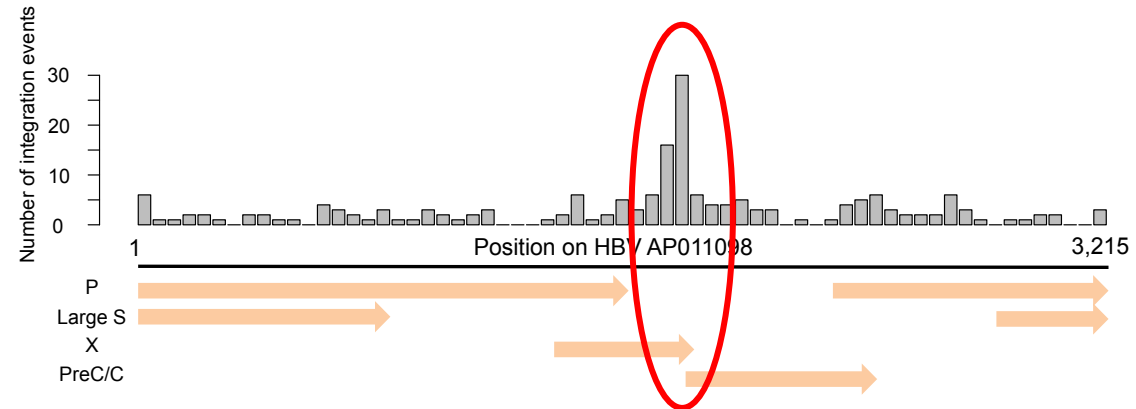
223 HBV integration events in 64 samples were identified.



HBV integration sites in human genome



HBV integration sites in HBV genome



Downstream of X gene

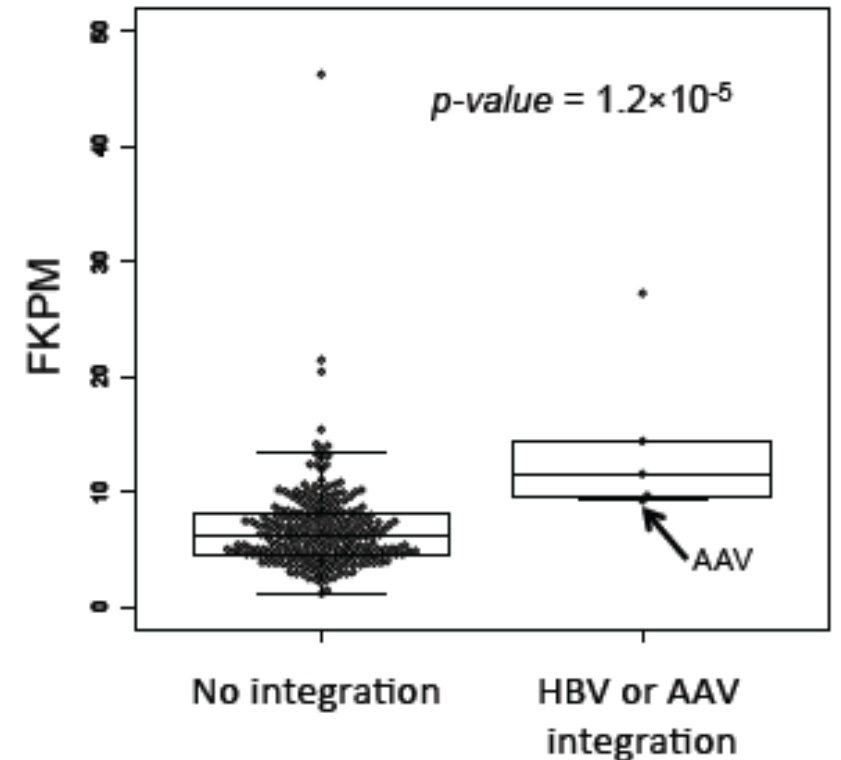
# Identification of adeno-associated virus (AAV)

- De novo* assembly of the unmapped reads from RNA-seq generated some long contigs that aligned to adeno-associated virus (AAV) in two liver cancer samples.

## Integrated genes

Sample	AAV genome		Human genome		Number of support read pair	Gene
	AAV sequence	Position	Chr	Position		
HX032_T01	NC_001401	4386	19	30304517	32	<i>CCNE1</i> intron
HX032_T01	NC_001401	4600	19	30304511	17	<i>CCNE1</i> intron
RK112_C01	NC_001401	3320	19	36213603	1	<i>MLL4</i> intron
RK112_C01	NC_001401	4438	19	36213955	1	<i>MLL4</i> intron
RK236_C01	NC_001401	2982	5	18813398	3	
RK236_C01	NC_001401	82	5	18813512	13	

## Expression level of *MLL4* gene

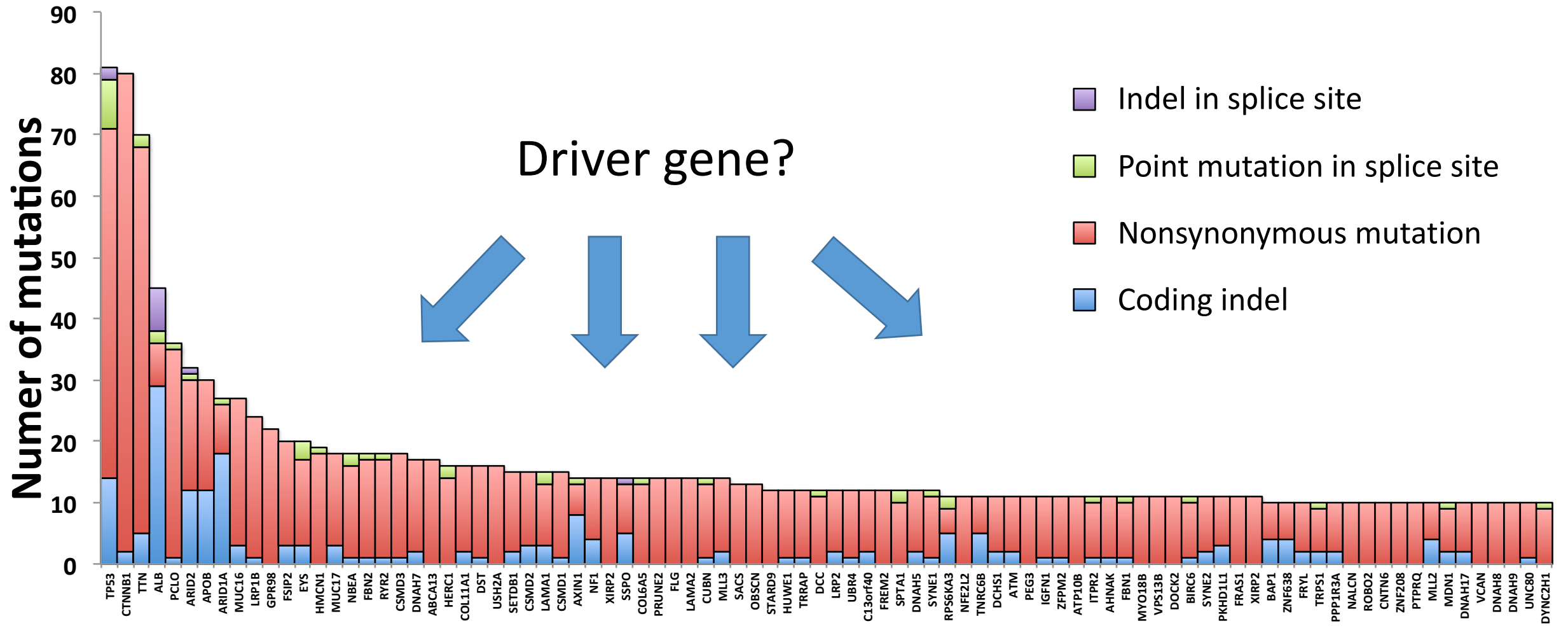


- Integration events of AAV into *MLL4* and *CCNE1* gene were identified. Expression level of *MLL4* gene in the integrated samples were higher than that of others. These results are consistent with a recent study (Nault *et al.* Nat Genet (2015)).

# Contents

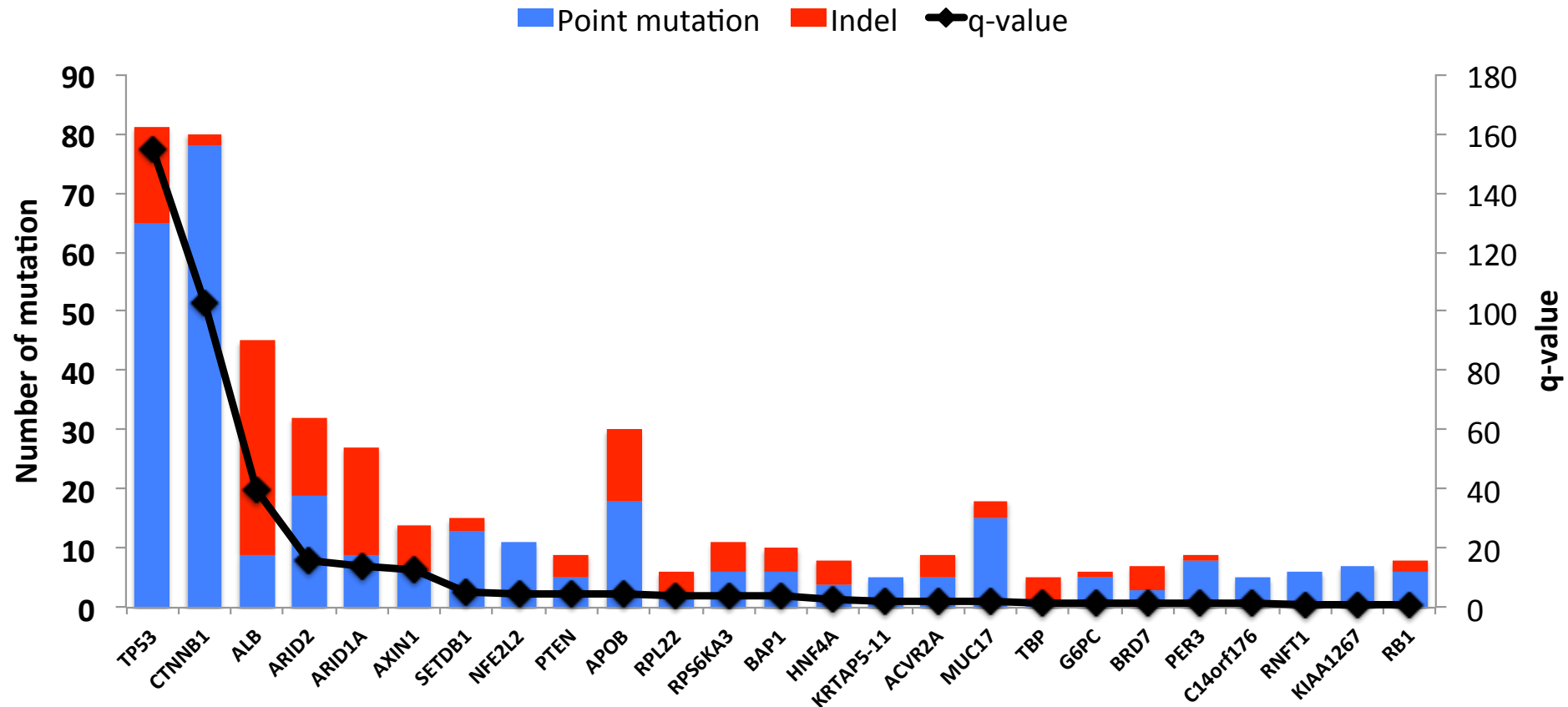
- Pattern of mutation
  - Point mutation and short indel
  - Structural variation
  - Integration of virus into human genome
- Driver genes and noncoding driver regions
  - Significantly mutated genes
  - Significantly mutated noncoding regions
- Structural variations
  - Structural variations
  - Structural variations and gene expression
  - Mutation and gene expression in *TERT*

# Protein alternating mutations (300 liver cancers)



# Identification of significantly mutated genes (1)

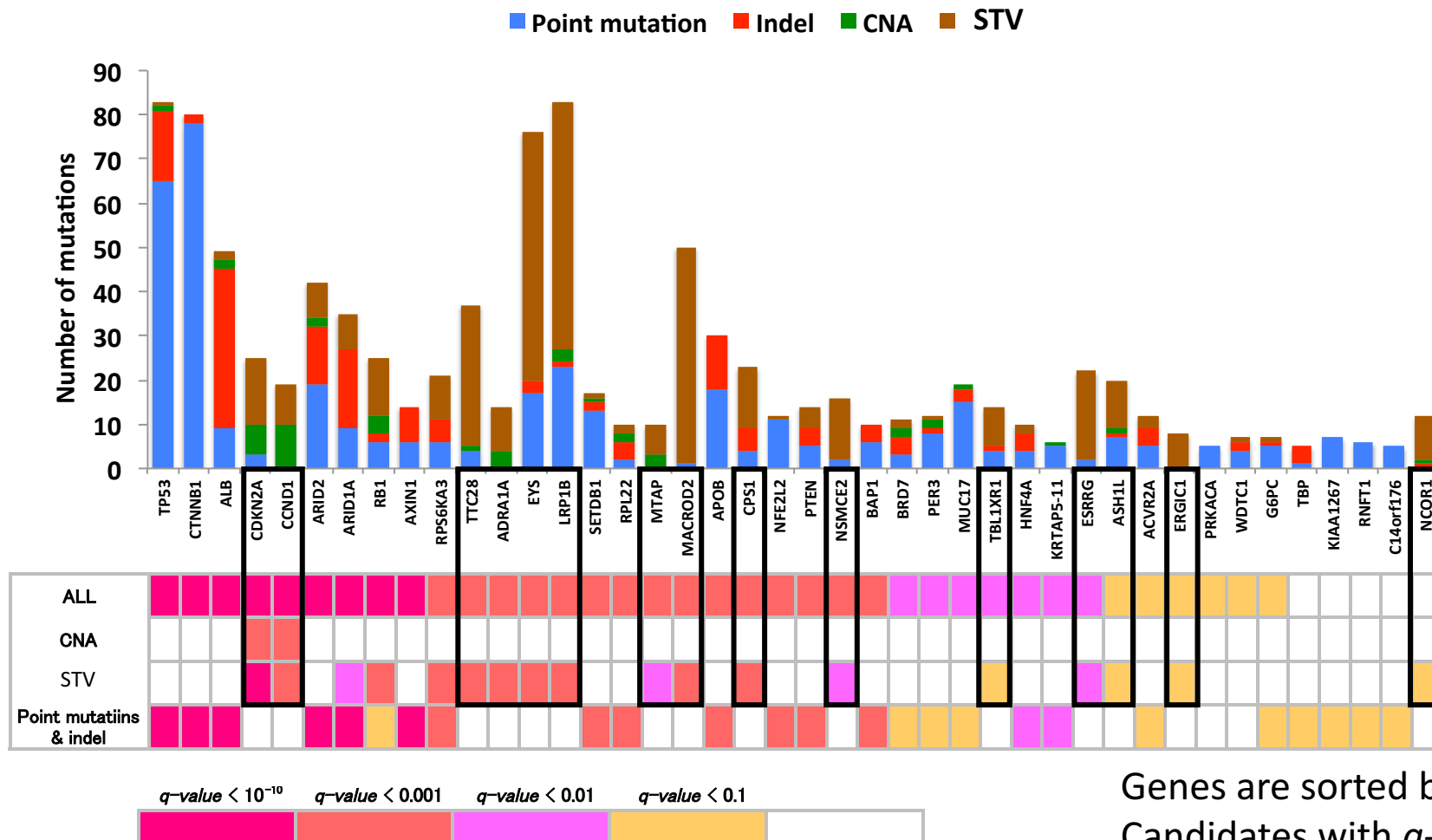
- (1) Estimate noncoding mutation rate within 1Mbp region
- (2) Calculate the expected number of mutations for each gene ((length) × (mutation rate))
- (3) Calculate probability that the observed number of mutation  $\geq$  expected number of mutations under the Poisson distribution



- Twenty-five genes, including *TP53*, *CTNNB1*, *ARID2*, *ARID1A*, *RB1*, *AXIN1*, *RPS6KA3*, *SETDB1*, *NFE2L2*, *BAP1*, and *HNF4A*, had a significantly large number of protein-altering mutations.

# Identification of significantly mutated genes (2)

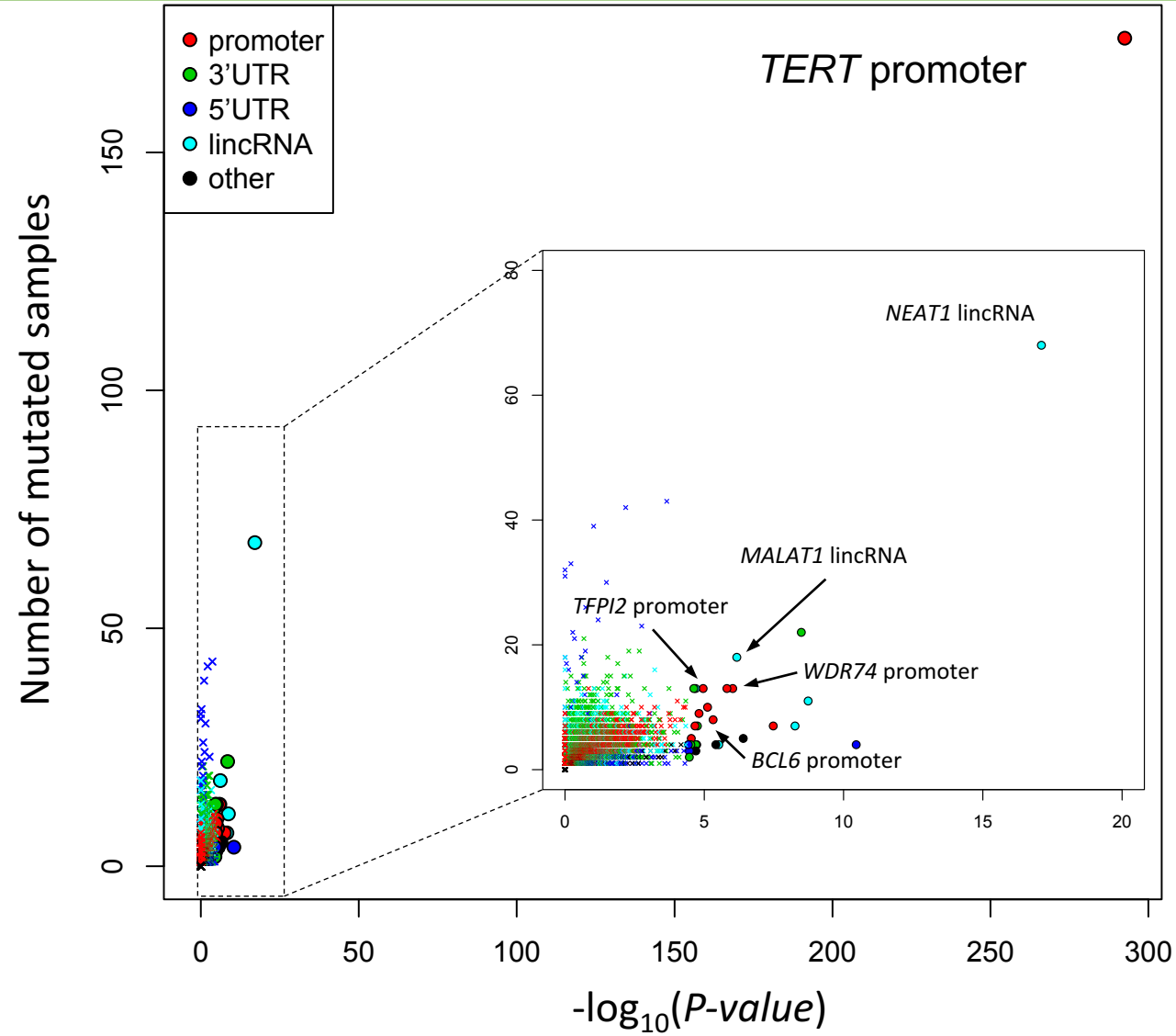
- We then combined the information on point mutations, indels, STVs, and CNAs and tested the significance of the number of mutations for each gene.
- Thirty-eight genes had a significantly large number of mutations.
- Genes with the terms ‘chromatin regulator’ and ‘regulation of cell cycle’ were significantly enriched.



Genes are sorted by  $q\text{-value}$ .  
Candidates with  $q\text{-value} < 0.1$  are shown.

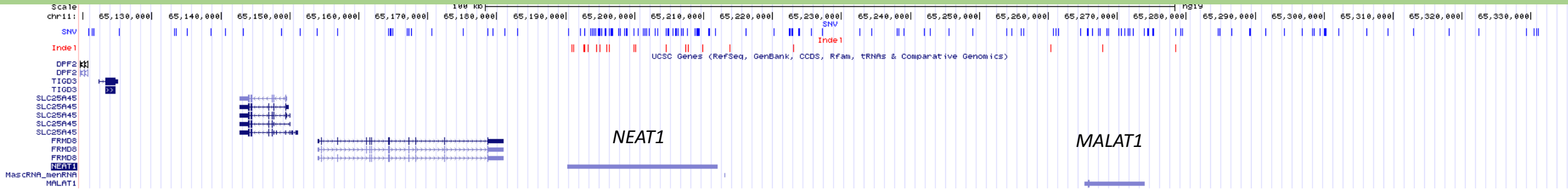


# Identification of significantly mutated noncoding elements (GENCODE annotation)



- In addition to the known *TERT* promoter, six long intergenic noncoding RNA (lincRNA) genes (*NEAT1* and *MALAT1*), ten promoter (*TFPI2*, *MED16*, and *WDR74*), and nine UTRs (*BCL6* and *AFF4*) were identified as regions with a significantly large number of mutations.

# lincRNA (*NEAT1* and *MALAT1*)



- ***NEAT1*** (Nuclear Enriched Abundant Transcript 1)

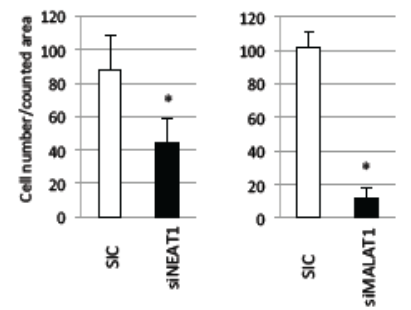
- NEAT1* has an essential role in constructing a subnuclear structure, paraspeckle<sup>1</sup>.
  - is differentially expressed in several cancers<sup>2,3,4,5</sup>
  - is induced by hypoxia and promote cell proliferation and invasion.
  - promotes cell proliferation and invasion through regulating miRNAs<sup>6,7,8,9,10</sup>.
  - is induced by TP53 and modulate replication stress response and chemosensitivity<sup>11</sup>.

- ***MALAT1*** (Metastasis-associated lung adenocarcinoma transcript 1)

- Expression level of *MALAT1* was associated with prognosis in liver cancer<sup>8</sup>.

- Knockdown of *NEAT1* or *MALAT1* in HepG2 cells decreased cell invasion

Invasion assay



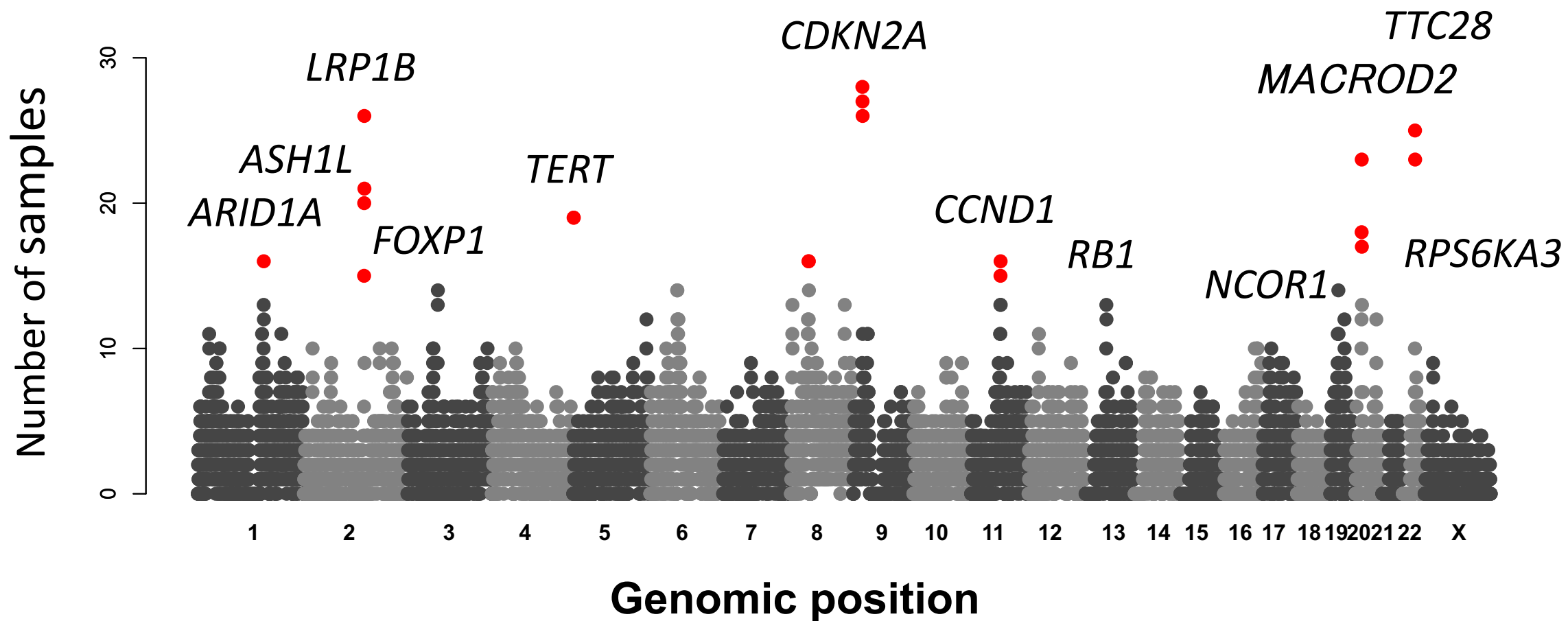
1; Hirose *et al.* Mol. Biol. Cell (2014), 2; Li *et al.* Oncotarget (2015), 3; Wu *et al.* Mol. Cancer (2016), 4; Chakravarty *et al.* Nat Comms (2014), 5 Choudhry *et al.* Oncogene (2015). 6; Wang *et al.* JECRC (2016), 7; Cao *et al.* Am J Cancer Res (2016), 8; Huang *et al.* BBRC (2016), 9; Ke *et al.* GRSB (2016), 10; Lai *et al.* Med Oncol (2012), 11 Adriaens *et al.* Net Med (2016)

# Contents

- Pattern of mutation
  - Point mutation and short indel
  - Structural variation
  - Integration of virus into human genome
- Driver genes and noncoding driver regions
  - Significantly mutated genes
  - Significantly mutated noncoding regions
- Structural variations
  - Structural variations
  - Structural variations and gene expression
  - Mutation and gene expression in *TERT*

# Distribution of structural variation (STV)

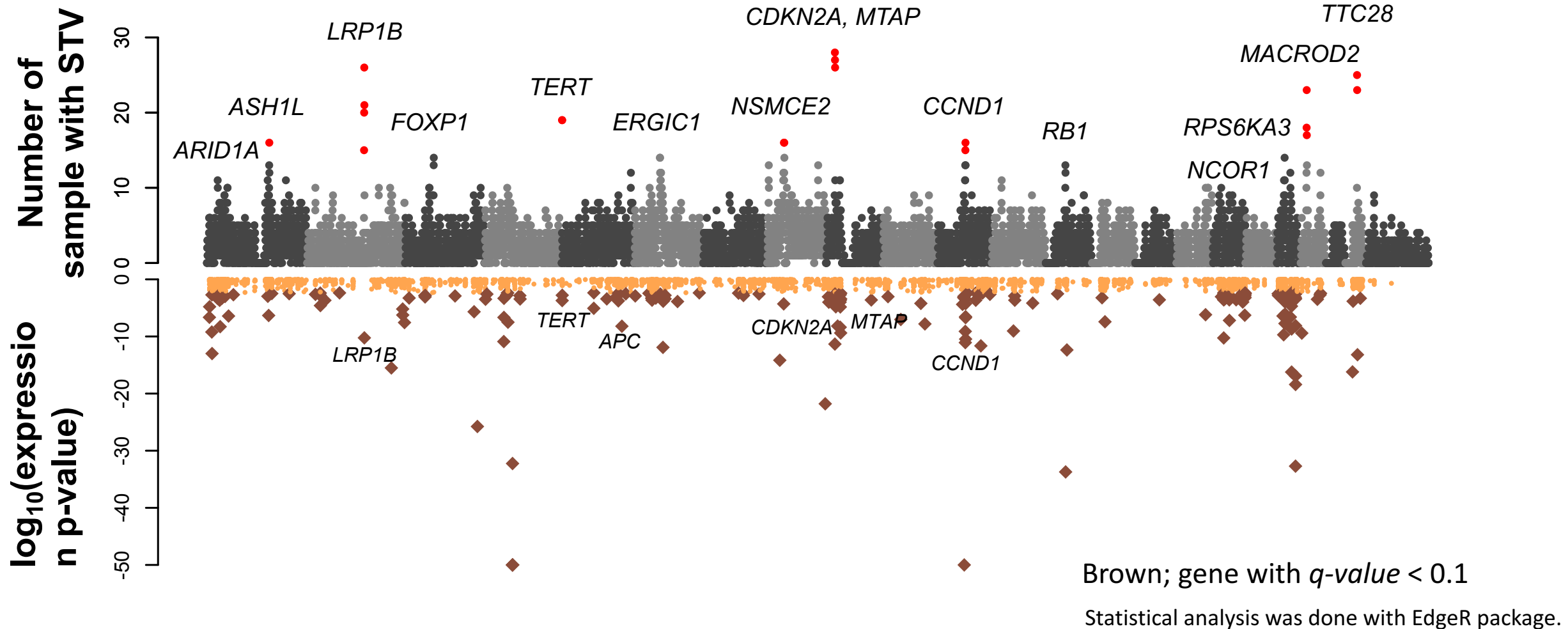
Number of samples with an STV breakpoint within 500-kb bins.



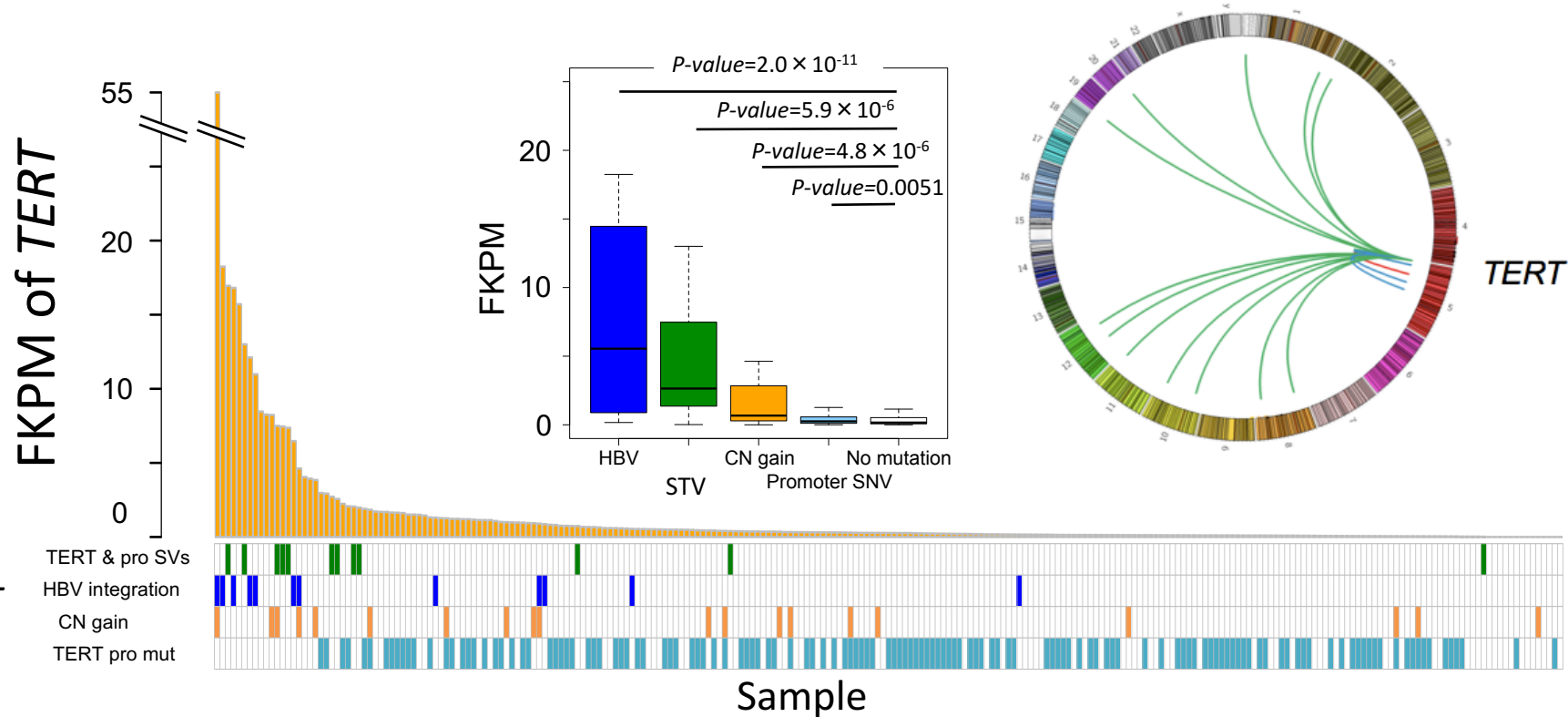
Red; Bins containing STV breakpoints in  $\geq 15$  samples (5%)

# STVs and gene expression

- We selected 4,940 genes that had STVs within 500 kb of their transcription start or end sites in  $\geq 6$  samples (2%) and compared the number of RNA-seq reads mapping to the genes for samples with and without STVs.
- Of these genes, 538 showed a significant difference in gene expression ( $q$  value  $< 0.1$ ).
- In the 538 genes, genes with terms of 'cell cycle' and 'mitotic cell cycle' were significantly enriched.

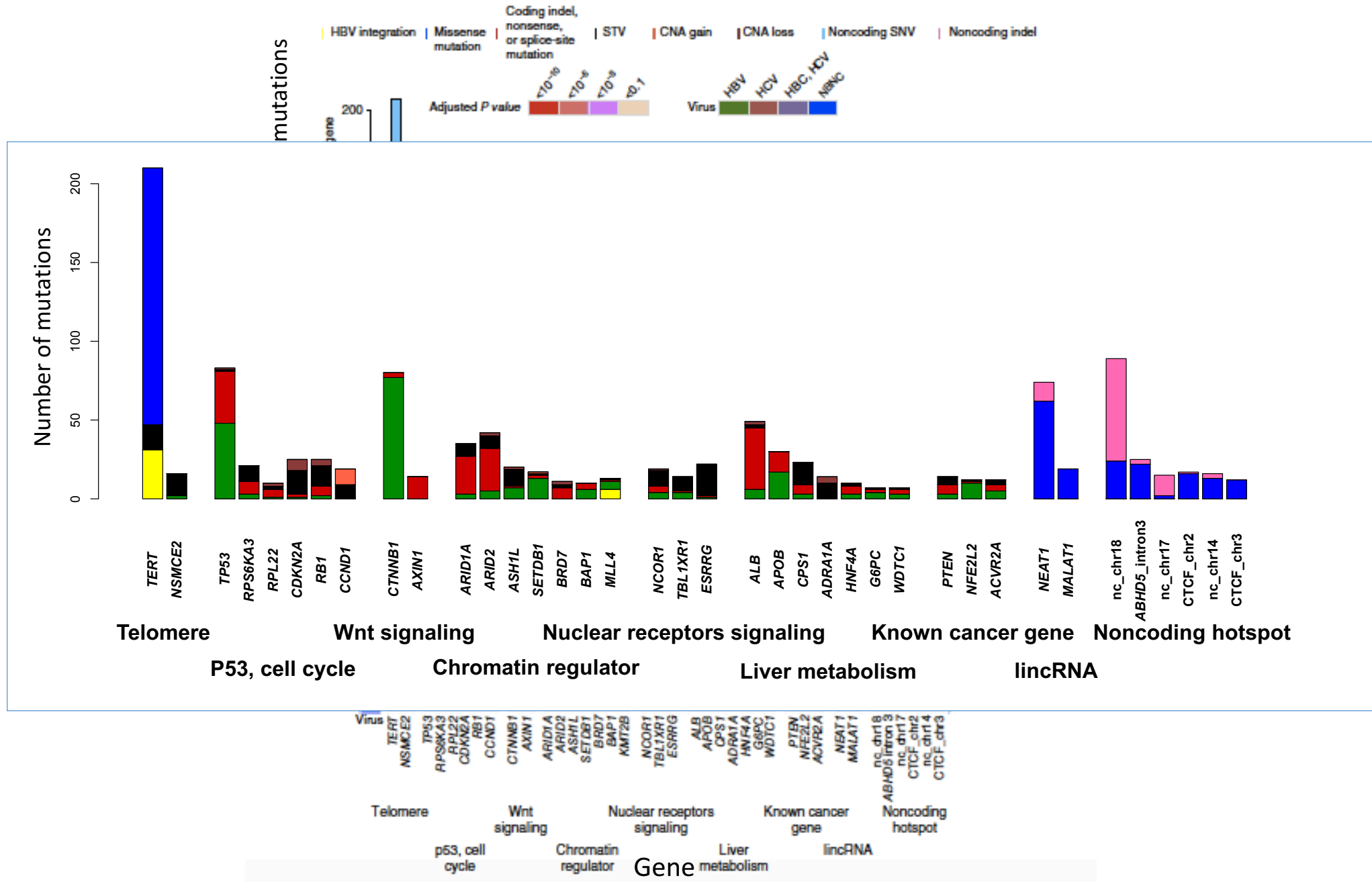


# Mutation in *TERT* gene and expression



- Promoter mutation (n=163), STVs (n=16), and HBV integrations (n=14) were identified in the *TERT* gene body and promoter region .
- Type of STVs were translocation and inversion.
- STVs, HBV and promoter point mutations were mutually exclusive.

# Summary of mutations in liver cancer



# Summary

- We analyzed whole-genome landscape of somatic alterations in 300 liver cancers.
- The number of point mutations were  $\sim 3/\text{Mbp}$ . But one sample (RK308) showed a hypermutated phenotype due to mismatch-repair deficiency.
- Our analysis identified 38 significantly mutated genes. Of these, chromatin regulators (*ASH1L*, and *SETDB1*), nuclear receptors (*TBL1XR1*, *NCOR1*, and *ESRRG*), DNA mismatch repair genes (*NSMCE2*, and *MACROD2*) and genes related to liver metabolism (*HNF4A*, and *G6PC*) were observed.
- Six long intergenic noncoding RNA (lincRNA) genes (*NEAT1* and *MALAT1*), ten promoters (*TFPI2*, *MED16*, and *WDR74*) and nine UTRs (*BCL6* and *AFF4*) were identified as regions with a significantly large number of mutations.
- A strong association between gene expression and STVs was observed.
- In the *TERT* regions, recurrent STVs and HBV integrations were detected, and associated with gene expression.



**YOKOZUNA**  
**(Grand Champ)**  
**横綱**



**TP53, CTNNB1, TERT**  
**(SNV, STV, HBV integration, amplification)**

**OH-ZEKI**  
**(Champ)**  
**大関**



**Wnt pathway genes**  
**AXIN1, APC...**  
**HBV integration**  
**lincRNA?**

**SEKI-WAKE**  
**関脇**



**Chromatin Regulators**  
**ARID2, ARID1A, MLL, MLL4,...**



**Many long-tail genes**  
**(幕内、十両)**

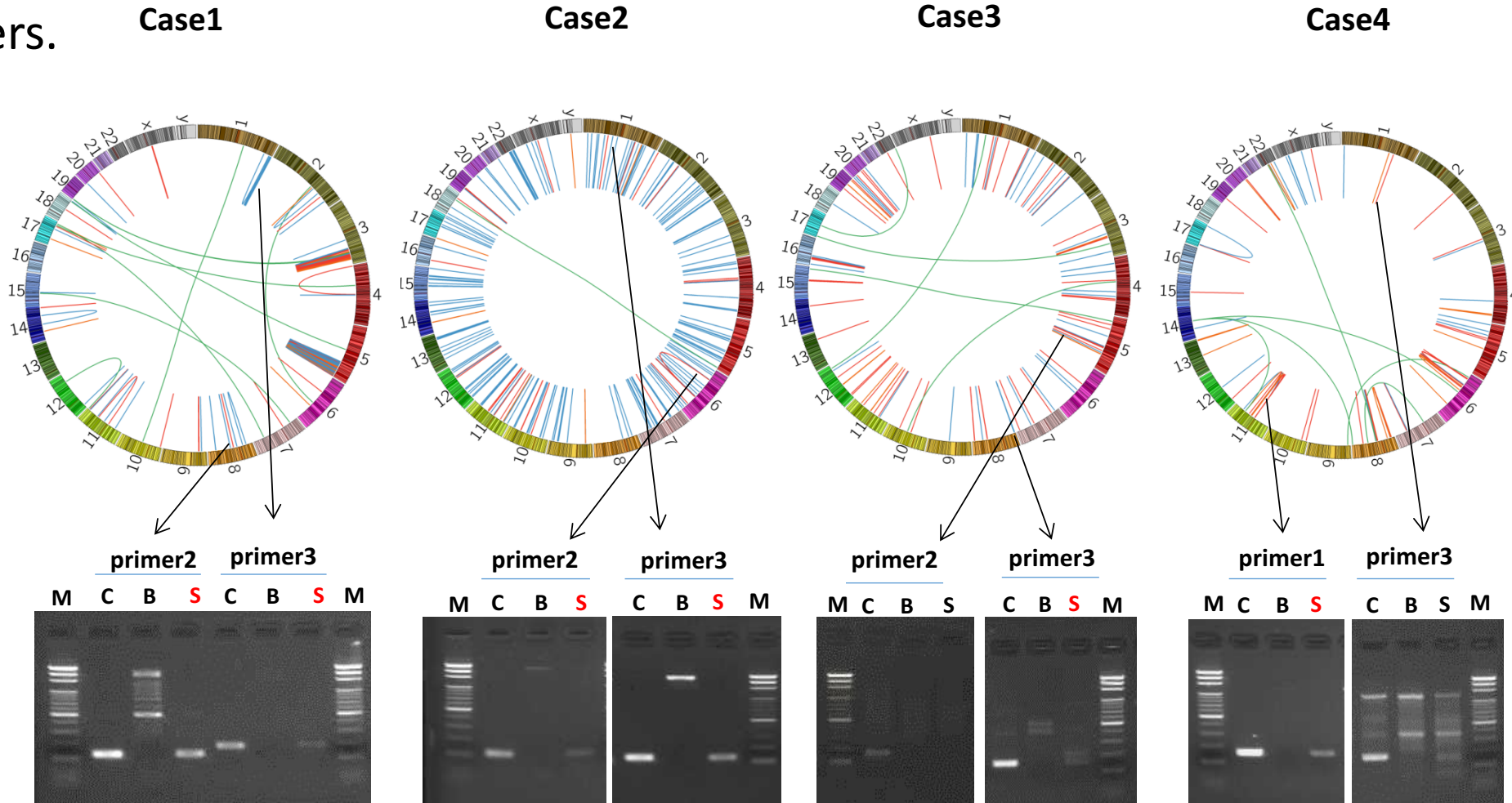
# Circulating tumor DNA analysis for liver cancers

Collaboration with Hiroshima University

Ono et al. **Cellular and Molecular Gastroenterology and Hepatology (2015)**

# “Personal” blood biomarkers to monitor the disease (1)

- Circulating tumor DNA (ctDNA) can be a good “personal” blood biomarker to monitor cancer.
- We detected ctDNA by structural variation.
- From WGS of 46 primary tumors, we selected STVs with high clonal proportion and designed PCR primers.



C: Cancer B: Blood Cell S: Serum M: Marker (100bp ladder)

# “Personal” blood biomarkers to monitor the disease (2)

- In the cfDNA from 46 patients, STV was detected by PCR in seven patients before surgery.
- The cumulative incidence of recurrence in the ctDNA-positive group was significantly worse than that in the ctDNA-negative group.

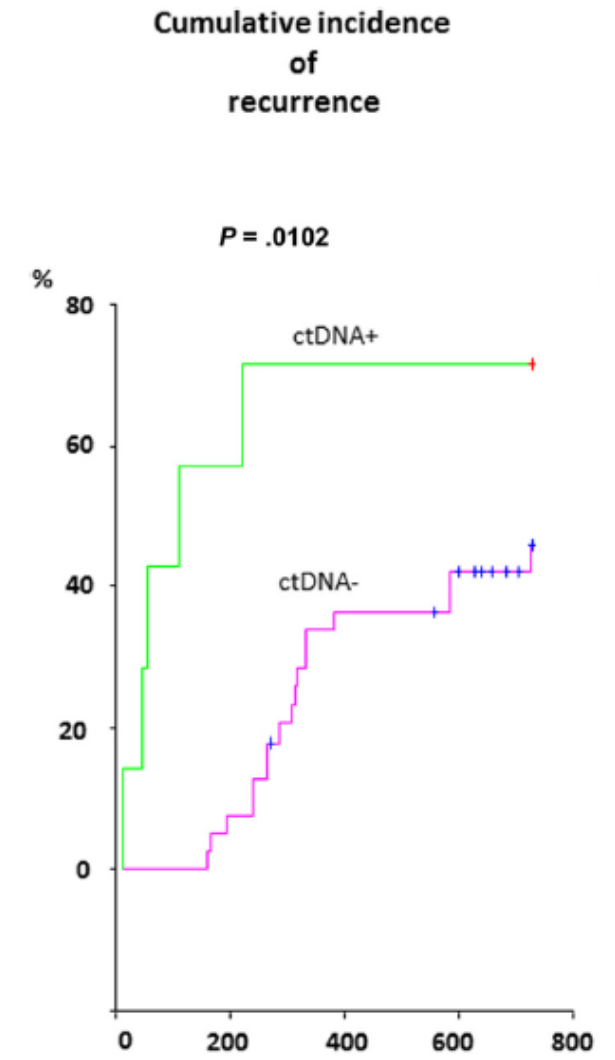
**Table 2.** Summary of Clinical Characteristics of Patients

Characteristic	Status of ctDNA Before Surgery	ctDNA Positive	ctDNA Negative	P Value
No. of patients	46	7	39	
Age at diagnosis (y)	67 (32–89)	68 (51–86)	67 (32–89)	.426
Gender (M/F)	35/11	5/2	30/9	.541 <sup>a</sup>
Etiology (HBV/HCV(SVR)/NBNC)	11/25 (6)/10	1/4(2)/2	10/21 (4)/8	.775 <sup>b</sup>
AFP (ng/mL) before surgery	60.5 (<0.5–57,410)	10,100 (12.7–57,410)	15.8 (<5–35330)	.004 <sup>c</sup>
DCP (mAU/mL) before surgery	57.5 (2.6–135,640)	23,156 (866–135,640)	37 (2.6–16123)	<.001 <sup>c</sup>
Tumor size (mm)	25.5 (10–140)	73 (35–140)	23 (10–125)	<.001 <sup>c</sup>

• **AFP and DCP levels and the rate of VP were significantly different between the cfDNA positive and negative samples.**

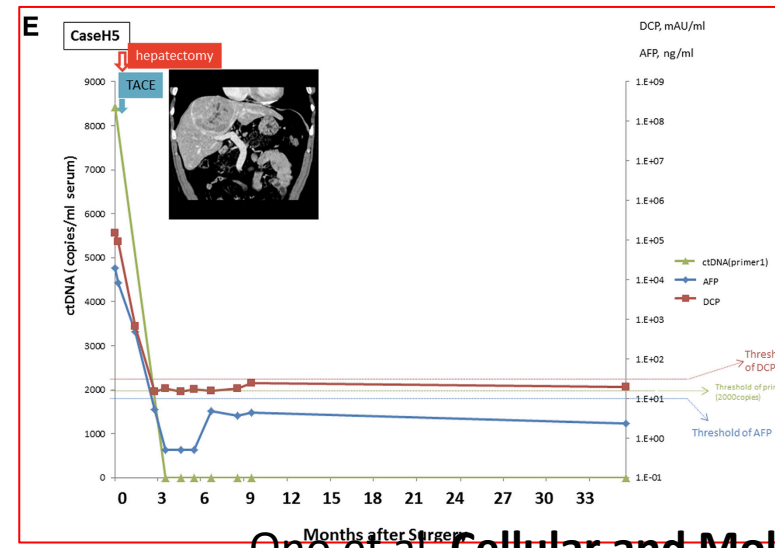
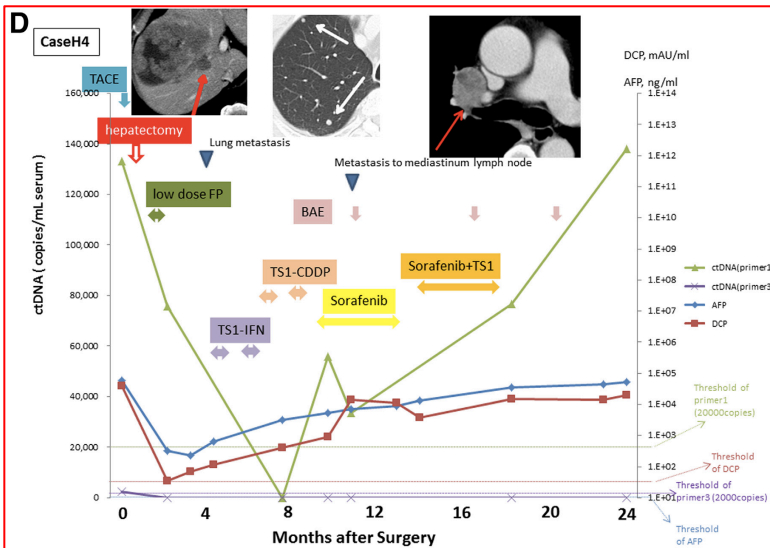
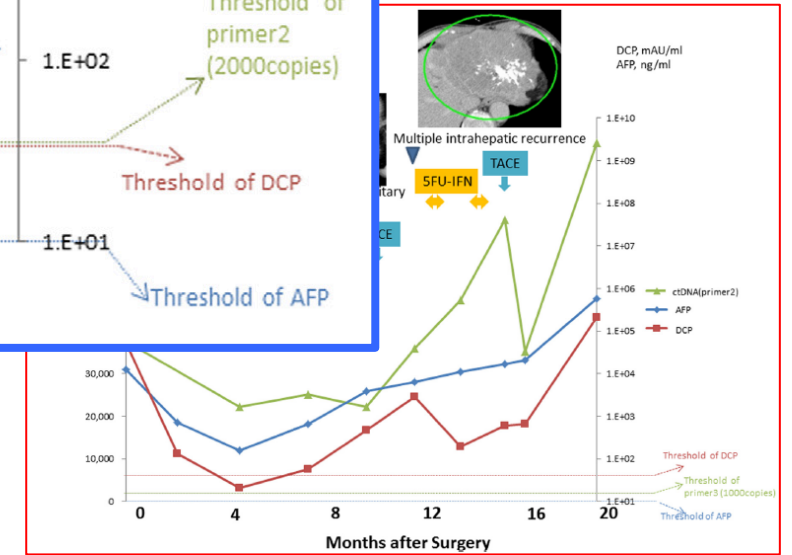
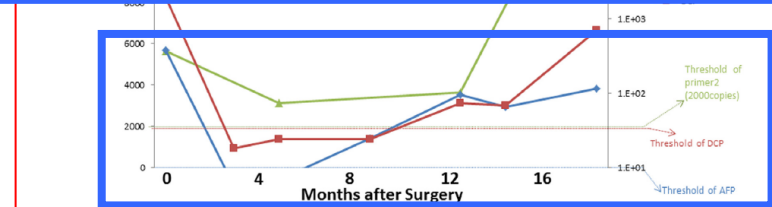
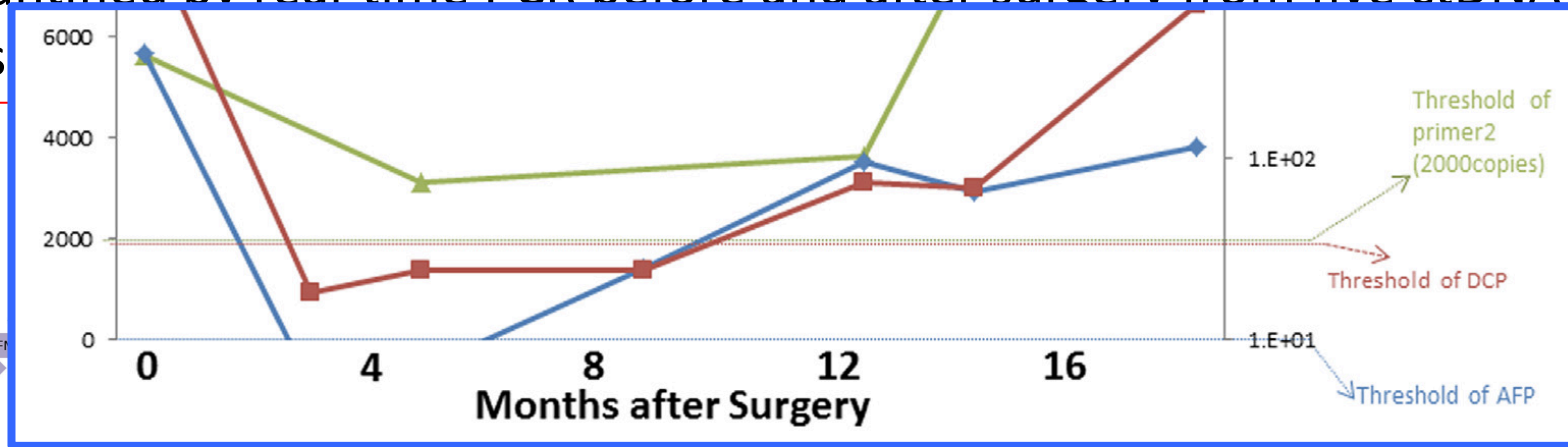
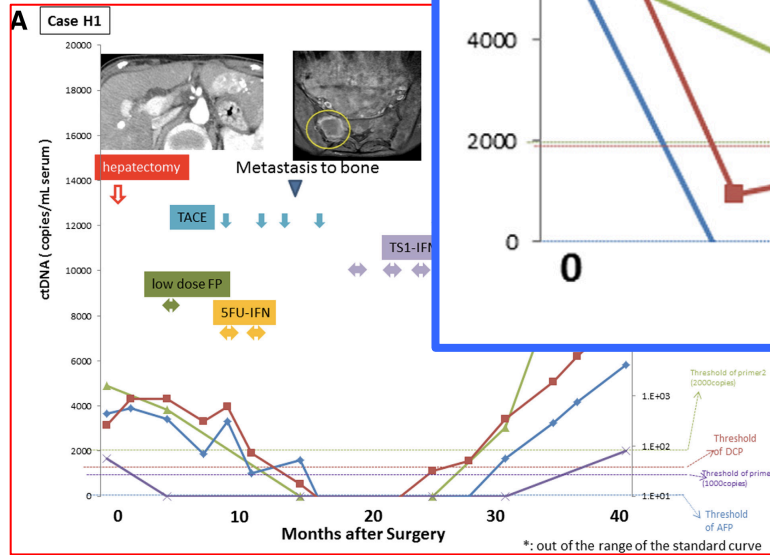
VV (0/1)	38/8	4/3	34/5	.089 <sup>a</sup>
VA (0/1)	45/1	6/1	39/0	.152 <sup>a</sup>

Note: AFP,  $\alpha$ -fetoprotein; DCP, des- $\gamma$ -carboxy prothrombin; HBV, hepatitis B virus; HCV (SVR), hepatitis C virus sustained virologic response; NBNC, neither HBsAg (+) nor anti-HCV (-); VA, hepatic artery; VP, microscopic vascular invasion to portal vein; VV, hepatic vein.  
<sup>a</sup>Fisher exact test.  
<sup>b</sup>Pearson chi-square test.  
<sup>c</sup>Mann-Whitney *U* test.



# Quantification of ctDNA in serum

- ctDNA was quantified by real-time PCR before and after surgery from five ctDNA positive samples.
- The time courses



- In a patient, AFP and DCP had been negative between 3 and 5 months after the surgery, but ctDNA was detected. ctDNA may be more sensitive than the conventional biomarkers in some patients.

# Collaborators

- **RIKEN**

**Mayuko Furuta, Tatsuhiko Tsunoda**, Hiroaki Taniguchi, Christopher P Wardell, Keith A Boroevich, Tetsuo Abe, Kaoru Nakano, Kazuhiro Maejima, Aya Sasaki-Oku, Ayako Ohsawa, Michiaki Kubo, **Hidewaki Nakagawa**

- **NCC**

**Yasushi Totoki, Mamoru Kato**, Hiromi Nakamura, Natsuko Hama, Fumie Hosoda, Yasuhito Arai, Shoko Ohashi, Nobuyoshi Hiraoka, Takuji Okusaka, Kazuaki Shimada, **Tatsuhiko Shibata**

- **University of Tokyo**

**Yuichi Shiraishi**, Hiroko Tanaka, Tetsuo Shibuya, Tomoko Urushidate, Genta Nagae, Shogo Yamamoto, Hiroki Ueda, Kenji Tatsuno, **Satoru Miyano, Hiroyuki Aburatani**

- **Hiroshima University School of Medicine**

**Atsushi Ono**, Yoshiiku Kawakami, Hiroshi Aikata, Koji Arihiro, Hideki Ohdan, **Kazuaki Chayama**

- **Hokkaido University Graduate School of Medicine**

Toru Nakamura, Satoshi Hirano

- **Keio University School of Medicine**

Hidenori Ojima

- **Wakayama Medical University**

Masaki Ueno, Shinya Hayami, Hiroki Yamaue

- **Osaka Medical Center for Cancer and Cardiovascular Diseases**

Kunihito Gotoh, Shigeru Marubashi, Shigeru Marubashi, Osamu Ishikawa

- **Tokyo Women's Medical University**

Shun-ichi Ariizumi, Masakazu Yamamoto



International  
Cancer Genome  
Consortium

